

## **Product Datasheet**

### **HiDi® DNA polymerase 9001**

**Product Name** HiDi® DNA polymerase

**Catalog Number** #9001

**Description** HiDi® stands for **H**igh **D**iscrimination of mismatches at the 3'-terminus of primers in PCR. This enzyme family is optimized for this feature and is the first choice for applications that rely on this property such as allele-specific PCR (asPCR), also known as allele-specific amplification (ASA).

Comparison studies with competitor products show that the HiDi® DNA polymerase family is the first choice for highly selective PCRs, such as genotyping by allele-specific PCR, HLA genotyping, analysis of single CpG methylation sites or the detection of mutations in a high background of wild-type sequences.

By using HiDi® DNA polymerase, less than 10 copies of a mutation can be detected in a background of >10,000 wild-type copies without any other tedious assay optimization.

**Applications:**

- Allele-specific PCR (asPCR), allele-specific amplification (ASA)
- HLA genotyping
- Analysis of single CpG methylation sites by PCR (methylation specific PCR, MSP)
- Mutation detection by PCR even in a high background of wild-type sequences
- Genotyping e.g., in CRISPR/Cas and TALEN approaches

This polymerase is also available as a **full-length Taq DNA polymerase** with a nuclease domain, featuring 100% compatibility with hydrolysis probes (TaqMan® probes etc.).

Several independently conducted studies show that HiDi® DNA polymerase is ideally suited for use in asPCR in numerous research areas ranging from mutation detection to genome editing. Please see "References" below.

*For research use and further manufacturing. Designed and manufactured under ISO13485*

**Tested Applications** End-Point, Real-Time

**Brand** myPOLS Biotec

**Storage** -20°C

## References

**Lifestyle-specific S-nitrosylation of protein cysteine thiols regulates Escherichia coli biofilm formation and resistance to oxidative stress.**

Barraud N, Létoffé S, Beloin C, Vinh J, Chiappetta G, Ghigo JM.NPJ Biofilms Microbiomes. 2021 Apr 13;7(1):34.

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**Primordial Germ Cell Migration and Histological and Molecular Characterization of Gonadal Differentiation in Pachón Cavefish *Astyanax mexicanus***

Imarazene B, Beille S, Jouanno E, Branthone A, Thermes V, Thomas M, Herpin A, Rétaux S, Guiguen Y Sex Dev. 2021 Mar 10;1-18.

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Miotto O, Sekihara M, Tachibana SI, Yamauchi M, Pearson RD, Amato R, Gonçalves S, Mehra S, Noviyanti R, Marfurt J, Auburn S, Price RN, Mueller I, Ikeda M, Mori T, Hirai M, Tavul L, Hetzel MW, Laman M, Barry AE, Ringwald P, Ohashi J, Hombhanje F, Kwiatkowski DP, Mita T. PLoS Pathog. 2020 Dec 15;16(12):e1009133.

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Yokouchi Y, Suzuki S, Ohtsuki N, Yamamoto K, Noguchi S, Soejima Y, Goto M, Ishioka K, Nakamura I, Suzuki S, Takenoshita S, Era T. Sci Rep. 2020 Aug 18;10(1):13927.

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**Development of sake yeast haploid set with diverse brewing properties using sake yeast strain Hiroshima no. 6 exhibiting sexual reproduction.**

Yamasaki R, Goshima T, Oba K, Kanai M, Ohdoi R, Hirata D, Akao T.

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Tsukumo Y, Naito M, Suzuki T.

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Link to publication:

<https://doi.org/10.1371/journal.pone.0229712>

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Biol Methods Protoc. 2020 Feb 10;5(1):bpaa004.

Link to publication:

<https://doi.org/10.1093/biomethods/bpaa004>

**Compensation of Disabled Organogeneses in Genetically Modified Pig Fetuses by Blastocyst Complementation.**

Matsunari H, Watanabe M, Hasegawa K, Uchikura A, Nakano K, Umeyama K, Masaki H, Hamanaka S, Yamaguchi T, Nagaya M, Nishinakamura R, Nakauchi H, Nagashima H.

Stem Cell Reports. 2020 Jan 14;14(1):21-33

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<https://doi.org/10.1016/j.stemcr.2019.11.008>

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Kikuchi Y, Hara-Kudo Y, Terajima J, Sugita-Konishi Y.  
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Nat Commun. 2019 Dec 6;10(1):5302

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**END-phenomenon negative bovine viral diarrhea virus that induces the host's innate immune response supports propagation of BVDVs with different immunological properties.**

Shiokawa M, Omatsu T, Katayama Y, Nishine K, Fujimoto Y, Uchiyama S, Kameyama KI, Nagai M, Mizutani T, Sakoda Y, Fukusho A, Aoki H.

Virology. 2019 Dec;538:97-110

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<https://doi.org/10.1016/j.virol.2019.09.016>

**Loss of ALBINO3b Insertase Results in Truncated Light-Harvesting Antenna in Diatoms.**

Nymark M, Volpe C, Hafskjold MCG, Kirst H, Serif M, Vadstein O, Bones AM, Melis A, Winge P.

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**Lhcx proteins provide photoprotection via thermal dissipation of absorbed light in the diatom *Phaeodactylum tricornutum*.**

Buck JM, Sherman J, Bártulos CR, Serif M, Halder M, Henkel J, Falciatore A, Lavaud J, Gorbunov MY, Kroth PG, Falkowski PG, Lepetit B.

Nat Commun. 2019 Sep 13;10(1):4167

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Sci Rep. 2019 Jul 9;9(1):9923

Link to publication:

<https://doi.org/10.1038/s41598-019-46357-8>

**A strategy to complement *PtaUREO1a* in TALEN knockout strains of *Phaeodactylum tricornutum***

S Madhuri, CR Bártulos, M Serif, B Lepetit, PG Kroth

Algal Research, 2019 May, 101469

Link to publication:

<https://doi.org/10.1016/j.algal.2019.101469>

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Nature. 2019 Feb;566(7742):110-114

Link to publication:

<https://doi.org/10.1038/s41586-019-0884-1>

**Precise and efficient nucleotide substitution near genomic nick via noncanonical homology-directed repair.**

Nakajima K, Zhou Y, Tomita A, Hirade Y, Gurumurthy CB, Nakada S.

Genome Res. 2018 Feb;28(2):223-230

Link to publication: <https://doi.org/10.1101/gr.226027.117>

**One-step generation of multiple gene knock-outs in the**

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**diatom *Phaeodactylum tricornutum* by DNA-free genome editing.**

Serif M, Dubois G, Finoux AL, Teste MA, Jallet D, Daboussi F.  
Nat Commun. 2018 Sep 25;9(1):3924

Link to publication:

<https://doi.org/10.1038/s41467-018-06378-9>

<b>Content</b>	S pack: 250 U, 5 U/ $\mu$ l, 1 x 50 $\mu$ l HiDi® DNA polymerase; 1 x 1.25 ml 10x HiDi reaction buffer
	M pack: 250 U, 5 U/ $\mu$ l, 1 x 200 $\mu$ l HiDi® DNA polymerase; 2 x 1.25 ml 10x HiDi reaction buffer