

Human CD264 / TRAIL R4 ELISA Kit

Instructions for use

Catalogue numbers:	1x96 tests:	850.850.096
	2x96 tests:	850.850.192

For research use only

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Table of Contents

1.	Intended Use	2
2.	Introduction	2
2.1.	Summary	2
2.2.	Principle of The Method	2
3.	Reagents Provided And Reconstitution	3
4.	Materials Required But Not Provided	3
5.	Storage Instructions	4
6.	Specimen Collection, Processing & Storage	4
7.	Safety & Precautions For Use	5
8.	Assay Preparation	6
8.1.	Assay Design	6
8.2.	Preparation of Wash Buffer	6
8.3.	Preparation of Standard Diluent Buffer 1X	6
8.4.	Preparation of Standard	7
8.5.	Preparation of Samples	7
8.6.	Preparation of Biotinylated Anti-CD264 / TRAIL R4	7
8.7.	Preparation of Streptavidin-HRP	8
9.	Method	9
10.	Data Analysis	10
11.	Assay Limitations	10
12.	Performance Characteristics	11
12.1.	Sensitivity	11
12.2.	Specificity	11
12.3.	Precision	11
12.4.	Dilution Parallelism	11
12.5.	Spike Recovery	11
12.6.	Expected Serum Values	11
13.	Bibliography	12
14.	Assay Summary	13

Human CD264 / TRAIL R4 ELISA KIT

1. Intended use

The Diaclone Human CD264 / TRAIL R4 ELISA kit is a solid phase sandwich ELISA for the *in-vitro* qualitative and quantitative determination of CD264 / TRAIL R4 / DcR2 in supernatants, buffered solutions or serum and plasma samples. This assay will recognise both natural and recombinant human CD264 / TRAIL R4.

This kit has been configured for research use only. Not suitable for use in therapeutic procedures.

2. Introduction

2.1. Summary

Human TRAIL R4, also called DcR2, CD264 or TRUNND is a type 1 TNF R family membrane protein which is a receptor for TRAIL. In the TRAIL receptor family, TRAIL R4 (DcR2) but also TRAIL R3 (DcR1) antagonize TRAIL-induced apoptosis whereas TRAIL R1 (DR4) and TRAIL R2 (DR5) transducer an apoptosis signal. TRAIL R4 has a truncated intracellular domain. Overexpression of TRAIL R4 has been shown to protect cells bearing TRAIL R1 and/or TRAIL R2 from Trail-induced apoptosis. Many tumor cell lines don't express the decoy receptors and are therefore sensitive to TRAIL if they express TRAIL R1 and/or TRAIL R2.

2.2. Principle of the method

A capture Antibody highly specific for CD264 / TRAIL R4 has been coated to the wells of the microtiter strip plate provided during manufacture. Binding of CD264 / TRAIL R4 in samples and known standards to the capture antibodies is completed and then any excess unbound analyte is removed.

During the next incubation period the binding of the Biotinylated anti-CD264 / TRAIL R4 secondary antibody to the analyte occurs. Any excess unbound secondary antibody is then removed.

The HRP conjugate solution is then added to every well including the zero wells, following incubation excess conjugate is removed by careful washing.

A chromogen substrate is added to the wells resulting in the progressive development of a blue coloured complex with the conjugate. The colour development is then stopped by the addition of acid turning the resultant final product yellow. The intensity of the produced coloured complex is directly proportional to the concentration of CD264 / TRAIL R4 present in the samples and standards.

The absorbance of the colour complex is then measured and the generated OD values for each standard are plotted against expected concentration forming a standard curve. This standard curve can then be used to accurately determine the concentration of CD264 / TRAIL R4 in any sample tested.

3. Reagents provided and reconstitution

Reagents (Store@2-8°C)	Quantity 1x96-well kit Cat no. 850.850.096	Quantity 2x96-well kit Cat no. 850.850.192	Reconstitution
Anti-CD264 / TRAIL R4 Coated Plate	1	2	Ready to use (96-well strip pre-coated plate)
Plastic plate covers	2	4	n/a
CD264 / TRAIL R4 Standard: 10 ng/ml	2	4	Reconstitute as directed on the vial (see Assay preparation, section 8)
Standard Diluent	1 (15ml)	1 (25ml)	10x concentrate, dilute in distilled water (see Assay preparation, section 8)
Biotinylated Anti-CD264 / TRAIL R4	1 (0.4ml)	2 (0.4ml)	Dilute in Biotinylated Antibody Diluent (see Assay preparation, section 8)
Biotinylated Antibody Diluent	1 (7ml)	1 (13ml)	Ready to use
Streptavidin-HRP	2 (5µl)	4 (5µl)	Add 0.5ml of Streptavidin-HRP Diluent prior to use (see Assay preparation, section 8)
Streptavidin-HRP Diluent	1 (12ml)	1 (23ml)	Ready to use
Wash Buffer	1 (10ml)	2 (10ml)	200x concentrate dilute in distilled water (see Assay preparation, section 8)
TMB Substrate	1 (11ml)	1 (24ml)	Ready to use
H ₂ SO ₄ Stop Reagent	1 (11ml)	2 (11ml)	Ready to use

4. Materials required but not provided

- Microtiter plate reader fitted with appropriate filters (450 nm required with optional 620 nm reference filter)
- Microtiter plate washer or wash bottle
- 10, 50, 100, 200 and 1,000µl adjustable single channel micropipettes with disposable tips
- 50-300µl multi-channel micropipette with disposable tips
- Multichannel micropipette reagent reservoirs
- Distilled water
- Vortex mixer
- Miscellaneous laboratory plastic and/or glass, if possible sterile

5. Storage Instructions

Store kit reagents between 2 and 8°C. Immediately after use remaining reagents should be returned to cold storage (2-8°C). Expiry of the kit and reagents is stated on box front labels. The expiry of the kit components can only be guaranteed if the components are stored properly, and if, in case of repeated use of one component, the reagent is not contaminated by the first handling.

Wash Buffer 1X: Once prepared, store at 2-8°C for up to 1 week.

Standard Diluent Buffer 1X: Once prepared, store at 2-8°C for up to 1 week.

Reconstituted Standard: Once prepared use immediately and do not store.

Diluted Biotinylated Anti-CD264 / TRAIL R4: Once prepared use immediately and do not store.

Diluted Streptavidin-HRP: Once prepared use immediately and do not store.

6. Specimen collection, processing & storage

Cell culture supernatants, human serum, plasma or other biological samples will be suitable for use in the assay. Remove serum from the clot or red cells, respectively, as soon as possible after clotting and separation.

Cell culture supernatants: Remove particulates and aggregates by spinning at approximately 1000 x g for 10 min.

Serum: Use pyrogen/endotoxin free collecting tubes. Serum should be removed rapidly and carefully from the red cells after clotting. Following clotting, centrifuge at approximately 1000 x g for 10 min and remove serum.

Plasma: EDTA, citrate and heparin plasma can be assayed. Spin samples at 1000 x g for 30 min to remove particulates. Harvest plasma.

Storage: If not analysed shortly after collection, samples should be aliquoted (250-500µl) to avoid repeated freeze-thaw cycles and stored frozen at -70°C. Avoid multiple freeze-thaw cycles of frozen specimens.

Recommendation: Do not thaw by heating at 37°C or 56°C. Thaw at room temperature and make sure that sample is completely thawed and homogeneous before use. When possible avoid use of badly haemolysed or lipemic sera. If large amounts of particles are present these should be removed prior to use by centrifugation or filtration.

7. Safety & precautions for use

- Handling of reagents, serum or plasma specimens should be in accordance with local safety procedures, e.g. CDC/NIH Health manual: "Biosafety in Microbiological and Biomedical Laboratories" 1984.
- Laboratory gloves should be worn at all times.
- Avoid any skin contact with H₂SO₄ and TMB. In case of contact, wash thoroughly with water.
- Do not eat, drink, smoke or apply cosmetics where kit reagents are used.
- Do not pipette by mouth.
- When not in use, kit components should be stored refrigerated as indicated on vials or bottles labels.
- All reagents should be warmed to room temperature before use. Lyophilized standards should be discarded after use.
- Once the desired number of strips has been removed, immediately reseal the bag to protect the remaining strips from deterioration.
- Cover or cap all reagents when not in use.
- Do not mix or interchange reagents between different lots.
- Do not use reagents beyond the expiration date of the kit.
- Use a clean disposable plastic pipette tip for each reagent, standard, or specimen addition in order to avoid cross contamination, for the dispensing of H₂SO₄ and TMB Substrate solutions, avoid pipettes with metal parts.
- Use a clean plastic container to prepare the washing solution.
- Thoroughly mix the reagents and samples before use by agitation or swirling.
- All residual washing liquid must be drained from the wells by efficient aspiration or by decantation followed by tapping the plate forcefully on absorbent paper. Never insert absorbent paper directly into the wells.
- The TMB Substrate solution is light sensitive. Avoid prolonged exposure to light. Also, avoid contact of the TMB Substrate solution with metal to prevent colour development. Warning TMB Substrate is toxic avoid direct contact with hands. Dispose off properly.
- If a dark blue colour develops within a few minutes after preparation, this indicates that the TMB Substrate solution has been contaminated and must be discarded. Read absorbances within 1 hour after completion of the assay.
- When pipetting reagents, maintain a consistent order of addition from well-to-well. This will ensure equal incubation times for all wells.
- Follow incubation times described in the assay procedure.
- Dispense the TMB Substrate within 15 min of the washing of the microtiter plate.

8. Assay Preparation

Bring all reagents to room temperature before use

8.1. Assay Design

Determine the number of microwell strips required to test the desired number of samples plus appropriate number of wells needed for running zeros and standards. Each sample, standard and zero should be tested **in duplicate**. Remove sufficient microwell strips for testing from the pouch immediately prior to use. Return any wells not required for this assay with desiccant to the pouch. Seal tightly and return to 2-8°C storage.

Example plate layout (example shown for a 6 point standard curve)

	Standards		Sample Wells									
	1	2	3	4	5	6	7	8	9	10	11	12
A	Std 1	Std 1										
B	Std 2	Std 2										
C	Std 3	Std 3										
D	Std 4	Std 4										
E	Std 5	Std 5										
F	Std 6	Std 6										
G	zero	zero										
H												

All remaining empty wells can be used to test samples in duplicate

8.2. Preparation of Wash Buffer

If crystals have formed in the concentrate Wash Buffer, warm it gently until complete dissolution.

Dilute the (200X) concentrate Wash Buffer 200 fold with distilled water to give a 1X working solution. Pour entire contents (10 ml) of the concentrate Wash Buffer into a clean 2,000 ml graduated cylinder. Bring final volume to 2,000 ml with glass-distilled or deionized water. Mix gently to avoid foaming. Transfer to a clean wash bottle and store at 2°-25°C.

8.3. Preparation of Standard Diluent Buffer 1X

If crystals have formed in the concentrate Standard Diluent, warm it gently until complete dissolution.

Dilute the (10X) concentrate Standard Diluent 10 fold with distilled water to give a 1X working solution. Pour entire contents of the concentrate Standard Diluent into a clean appropriate graduated cylinder. Bring to final volume with glass-distilled or deionized water. Transfer to a clean wash bottle and store at 2°-25°C. Please see example volumes below:

Standard Diluent concentrate (ml)	Distilled water (ml)
15	135
25	225

8.4. Preparation of Standard

Standard vials must be reconstituted with the volume of Standard Diluent Buffer 1X shown on the vial immediately prior to use. This reconstitution gives a stock solution of 10 ng/ml of CD264 / TRAIL R4. Mix the reconstituted standard gently by inversion only. Serial dilutions of the standard are made directly in the assay plate to provide the concentration range from 10 to 0.312 ng/ml. A fresh standard curve should be produced for each new assay.

- Immediately after reconstitution add 200µl of the reconstituted standard to wells A1 and A2, which provides the highest concentration standard at 10 ng/ml.
- Add 100µl of Standard Diluent Buffer 1X to the remaining standard wells B1 and B2 to F1 and F2.
- Transfer 100µl from wells A1 and A2 to B1 and B2. Mix the well contents by repeated aspirations and ejections taking care not to scratch the inner surface of the wells.
- Continue this 1:1 dilution using 100µl from wells B1 and B2 through to wells F1 and F2 providing a serial diluted standard curve ranging from 10 ng/ml to 0.312 ng/ml.
- Discard 100µl from the final wells of the standard curve (F1 and F2).

Alternatively these dilutions can be performed in separate clean tubes and immediately transferred into the relevant wells.

8.5. Preparation of Samples

Before testing, human serum or plasma samples have to be diluted 1:4 in Standard Diluent Buffer 1X.

8.6. Preparation of Biotinylated Anti-CD264 / TRAIL R4

It is recommended this reagent is prepared immediately before use. Dilute the Biotinylated Anti-CD264 / TRAIL R4 with the Biotinylated Antibody Diluent in an appropriate clean glass vial using volumes appropriate to the number of required wells. Please see example volumes below:

Number of wells required	Biotinylated Antibody (µl)	Biotinylated Antibody Diluent (µl)
16	40	1060
24	60	1590
32	80	2120
48	120	3180
96	240	6360

8.7. Preparation of Streptavidin-HRP

It is recommended to centrifuge vial for a few seconds in a microcentrifuge to collect all the volume at the bottom.

Dilute the 5 μ l vial with 0.5ml of Streptavidin-HRP Diluent **immediately before use**. Do not keep this diluted vial for future experiments. Further dilute the HRP solution to volumes appropriate for the number of required wells in a clean glass vial. Please see example volumes below:

Number of wells required	Streptavidin-HRP (μ l)	Streptavidin-HRP Diluent (ml)
16	30	2
24	45	3
32	60	4
48	75	5
96	150	10

9. Method

We strongly recommend that every vial is mixed thoroughly without foaming prior to use.

Prepare all reagents as shown in section 8.

Note: final preparation of Biotinylated Antibody (section 8.6) and Streptavidin-HRP (section 8.7) should occur immediately before use.

Assay Step		Details
1.	Addition	Prepare standard curve as shown in section 8.4 above and add in duplicate to appropriate wells
2.	Addition	Add 100µl of each Sample and zero (Standard Diluent Buffer 1X) in duplicate to appropriate number of wells
3.	Incubation	Cover with a plastic plate cover and incubate at room temperature (18 to 25°C) for 2 hours
4.	Wash	Remove the cover and wash the plate as follows: a) Aspirate the liquid from each well b) Dispense 0.3 ml of 1x Wash Buffer into each well c) Aspirate the contents of each well d) Repeat step b and c another two times
5.	Addition	Add 50µl of diluted Biotinylated Anti-CD264 / TRAIL R4 to all wells
6.	Incubation	Cover with a plastic plate cover and incubate at room temperature (18 to 25°C) for 1 hour
7.	Wash	Repeat wash step 4.
8.	Addition	Add 100µl of diluted Streptavidin-HRP solution into all wells
9.	Incubation	Cover with a plastic plate cover and incubate at room temperature (18 to 25°C) for 30 min
10.	Wash	Repeat wash step 4.
11.	Addition	Add 100µl of ready-to-use TMB Substrate solution into all wells
12.	Incubation	Incubate in the dark for 10-20 minutes* at room temperature. Avoid direct exposure to light by wrapping the plate in aluminium foil.
13.	Addition	Add 100µl of H₂SO₄ Stop Reagent into all wells
Read the absorbance value of each well (immediately after step 13.) on a spectrophotometer using 450 nm as the primary wavelength and optionally 620 nm as the reference wave length (610 nm to 650 nm is acceptable).		

**Incubation time of the TMB Substrate is usually determined by the ELISA reader performance. Many ELISA readers only record absorbance up to 2.0 O.D. Therefore the colour development within individual microwells must be observed by the analyst, and the substrate reaction stopped before positive wells are no longer within recordable range.*

10. Data Analysis

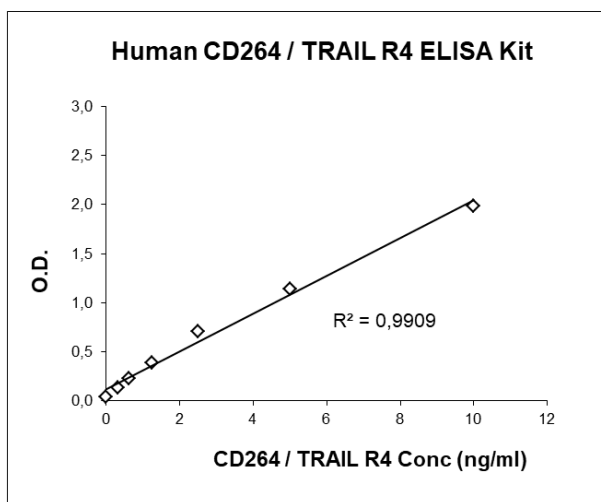
Calculate the average absorbance values for each set of duplicate standards and samples. Ideally duplicates should be within 20% of the mean.

Generate a linear standard curve by plotting the average absorbance of each standard on the vertical axis versus the corresponding CD264 / TRAIL R4 standard concentration on the horizontal axis.

The amount of CD264 / TRAIL R4 in each sample is determined by extrapolating OD values against CD264 / TRAIL R4 standard concentrations using the standard curve.

Example CD264 / TRAIL R4 Standard curve

Standard	CD264 / TRAIL R4 Conc (ng/ml)	OD (450nm) mean	CV (%)
1	10	1.989	0.6
2	5	1.142	0.7
3	2.5	0.705	7.3
4	1.25	0.381	1.3
5	0.625	0.224	1.7
6	0.312	0.134	9.5
zero	0	0.039	-



Note: curve shown above should not be used to determine results. Every laboratory must produce a standard curve for each set of microwell strips assayed.

For samples human serum or plasmas which have been diluted 1:4 according to the protocol, the calculated concentration should be multiplied by the dilution factor (x4).

11. Assay limitations

Do not extrapolate the standard curve beyond the maximum standard curve point. The dose-response is non-linear in this region and good accuracy is difficult to obtain. Concentrated samples above the maximum standard concentration must be diluted with Standard Diluent Buffer or with your own sample buffer to produce an OD value within the range of the standard curve. Following analysis of such samples always multiply results by the appropriate dilution factor to produce actual final concentration.

The influence of various drugs on end results has not been investigated. Bacterial or fungal contamination and laboratory cross-contamination may also cause irregular results.

Improper or insufficient washing at any stage of the procedure will result in either false positive or false negative results. Completely empty wells before dispensing fresh Wash Buffer, fill with Wash Buffer as indicated for each wash cycle and do not allow wells to sit uncovered or dry for extended periods.

Disposable pipette tips, flasks or glassware are preferred, reusable glassware must be washed and thoroughly rinsed of all detergents before use.

As with most biological assays conditions may vary from assay to assay therefore **a fresh standard curve must be prepared and run for every assay.**

12. Performance Characteristics

12.1. Sensitivity

The sensitivity or minimum detectable dose of CD264 / TRAIL R4 using this Diaclone Human CD264 / TRAIL R4 ELISA kit was found to be **64 pg/ml**. This was determined by adding 3 standard deviations to the mean OD obtained when the zero standard was assayed 32 times.

12.2. Specificity

The assay recognizes both natural and recombinant human CD264 / TRAIL R4. To define the specificity of this ELISA several proteins were tested for cross reactivity. There was no cross reactivity observed for any protein tested: TRAIL, CD117, IL-6R, IL-2R, CD116, TRAIL R2, TRAIL R3, TRAIL R1, CD178 and Granzyme B.

12.3. Precision

Intra-assay					Inter-assay				
Sample	n	Mean (pg/mL)	SD	CV%	Sample	n	Mean (pg/mL)	SD	CV%
A	8	5723	212	3.7	A	12	6531	627	9.6
B	8	3063	218	7.1	B	12	3123	369	11.8

12.4. Dilution Parallelism

One human serum containing 10 ng/ml of CD264 / TRAIL R4 was serially diluted in standard buffer diluent. Linear regression of samples versus the expected concentration yielded a correlation coefficient of 0.99.

12.5. Spike Recovery

The spike recovery was evaluated by spiking concentrations of natural CD264 / TRAIL R4 in human serum. Recoveries ranged from 68% to 87% with an overall **mean recovery of 77%**.

12.6. Expected serum values

CD264 / TRAIL R4 is not detected in healthy donor sera (95 sera tested).

13. Bibliography

1. Degli-Esposti MA and al, Immunity 1997, 7: 813-20.
2. Golstein P, Curr Biol. 1997 Dec 1;7(12):R750-3.
3. Kimberley FC and al, Cell Res. 2004 Oct;14(5):359-72.
4. Marsters SA and al, Curr Biol. 1997, 7(12):1003-6.
5. Pan G and al, FEBS Lett. 1998 Mar 6;424(1-2):41-5.

14. Assay Summary

Total procedure length : 3h45min

Add 100µl of diluted Samples and diluted Standards



Incubate 2 hours at room temperature



Wash three times



Add 50µl of diluted Biotinylated Antibody



Incubate 1 hour at room temperature



Wash three times



Add 100µl of diluted Streptavidin-HRP



Incubate 30 min at room temperature



Wash three times



Add 100µl of TMB Substrate
Protect from light. Let the color develop for 10-20 min.



Add 100µl of Stop Reagent



Read Absorbance at 450 nm

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